

ADMINISTRATION OF QUEUINE TO MICE RELIEVES MODIFIED NUCLEOSIDE
QUEUOSINE DEFICIENCY IN EHRLICH ASCITES TUMOR tRNA

Jon R. Katze

Department of Microbiology and Immunology, University of Tennessee
Center for the Health Sciences
Memphis, Tennessee 38163

and

William T. Beck

Division of Biochemical and Clinical Pharmacology
St. Jude Children's Research Hospital
Memphis, Tennessee 38101

Received July 29, 1980

SUMMARY: The tRNA from Ehrlich ascites tumor cells is deficient in the modified nucleoside Q (queuosine). Continuous infusion of Q base (queuine) to tumor-bearing mice reverses the deficiency of Q in Ehrlich ascites tRNA, and coincidentally, causes an inhibition of tumor growth.

The modified nucleoside Q (queuosine), 7-[[cis-4,5-dihydroxy-2-cyclopenten-1-yl)-amino]-methyl]-7-deazaguanosine, occurs exclusively in the first position of the anticodons of tRNA^{Tyr}, tRNA^{His}, tRNA^{Asn} and tRNA^{ASP} [these tRNAs form a set in that they accommodate the codons NAC^{II} (where N=U,C, A, or G)] (1). The occurrence of Q in the anticodon suggests the possibility of Q-dependent translational control; however, functional differences between Q-free tRNA ([Q-]tRNA) and Q-containing tRNA ([Q+]tRNA) have not been detected using protein synthesis systems *in vitro* (2-5). Unlike other modifications of tRNA, Q is formed in part as the free base and then is incorporated at the posttranscriptional level into tRNA by the enzyme tRNA-guanine transglycosylase (6,7) (Fig. 1). The mammalian enzyme exchanges queuine (the base of Q) for guanine *in vitro* (6,7). In contrast, the bacterial enzyme exchanges a precursor of queuine into tRNA, and further modification to yield Q occurs at the polynucleotide level (7). The mammalian reaction presented in Fig. 1 is

presumably physiologically significant, because a factor in bovine amniotic fluid that enables cultured mammalian cells to synthesize [Q+]tRNA has been identified as queuine (8). Normal mammalian tRNA of the NA_C^U codon set is almost completely substituted with Q or hexose-containing derivatives of Q (9,10); however, Q-deficient tRNA has been observed in (a) certain tumors (2,9-12), (b) fetal and regenerating liver (8,13,14), (c) reticulocytes (4,13,14), and (d) germ-free mice fed a defined diet (16). The present study was designed to determine if the administration of queuine could relieve the deficiency of Q in tumor tRNA in vivo. The results demonstrate that the administration of queuine to tumor-bearing mice reverses the deficiency of Q in Ehrlich ascites tRNA and, coincidentally, causes an inhibition of the growth of these neoplastic cells.

MATERIALS AND METHODS

Queuine was isolated from bovine amniotic fluid (7- to 9-month gestation, Irvine Scientific Co.) by a modification (J.R. Katze, unpublished) of a described procedure (6). Animals were housed in cages containing hardwood chip bedding and were fed standard laboratory chow (Ralston Purina). tRNA was isolated, aminoacylated using a crude aminoacyl-tRNA synthetase preparation from mouse liver, and recovered by DEAE-cellulose chromatography after aminoacylation as described (10). Reversed-phase chromatography employed a column (0.6 x 21 cm) of RPC-5 support developed at 27°C with a 300 ml 0.48 M - 1.0 M NaCl linear gradient in standard buffer, with 1.5 ml fractions collected, and the radioactivity measured as described (10) [3H]His-tRNA was oxidized by periodate as described (16), and then recovered from the reaction by absorption and elution from DEAE-cellulose prior to RPC-5 chromatography.

RESULTS AND DISCUSSION

The data shown in Figure 2 and Table 1 demonstrate that Ehrlich ascites tumor tRNA^{His} is significantly Q-deficient, confirming and expanding other observations (9). The deficiency of Q in tRNA^{ASP} in Ehrlich ascites tumor cells is negligible (data not shown), in keeping with evidence that tRNA^{ASP} species are preferred substrates for tRNA-guanine transglycosylase (ref. 17 and W.R. Farkas and J.R. Katze, unpublished). The side chain of Q contains a cis-diol group which, when oxidized by periodate (1), causes a specific delay in the elution of [Q+]tRNAs from RPC-5 columns (18). The

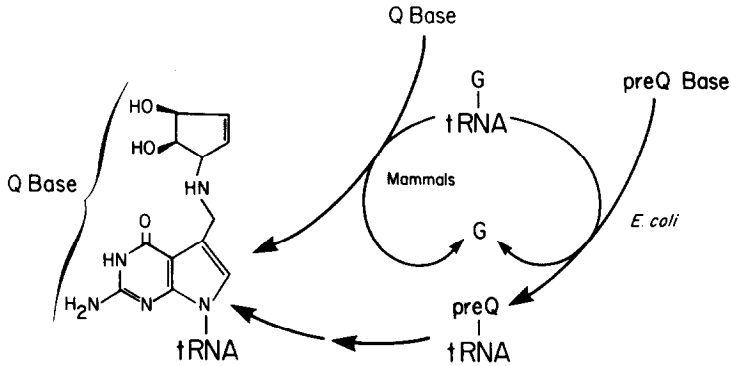


Fig. 1. Comparison of tRNA-Guanine Transglycosylase Catalyzed Steps of Q Synthesis in Mammals and *E. coli*. The abbreviation preQ base represents 7-(aminomethyl)-7-deazaquinoline (see ref. 7). Whether the bacterial pathway also operates in mammals is unknown.

chromatogram in Fig. 2 shows a representative sample of tumor tRNA^{His} before and after treatment with periodate. Only tRNA^{His}₁ was affected by periodate and is thus identified as [Q+]; by contrast, tRNA^{His}₂ and tRNA^{His}₃ were unaffected by periodate and are [Q-]. Mammalian tRNA^{His}₁ and tRNA^{His}₂ were shown

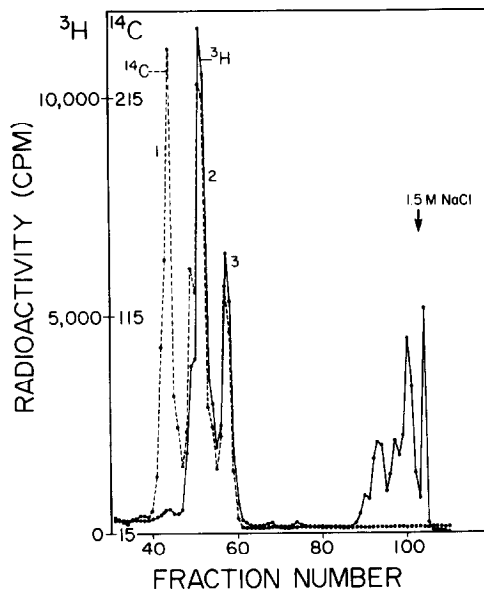


Fig. 2. Co-Chromatographic comparison of Ehrlich ascites tumor tRNA^{His} before and after periodate oxidation. [¹⁴C]His-tRNA is from the tumor of one mouse (Table 1, Exp. 2, untreated with queuine). [³H]His-tRNA is from the same tRNA sample, but treated with periodate after aminoacylation. Additional experimental details are found in Materials and Methods.

TABLE 1. Effect of Queuine on tRNA^{His} Isoaccepting Species From Ehrlich Ascites Tumors*

Group	N	Tumor tRNA ^{His}			Liver tRNA ^{His}		
		%1 [O+]	%2 [O-]	%3 [O-]	%1 [O+]	%2 [O-]	%3 [O-]
Tumor-bearing	3	46 ± 14	42 ± 13	12 ± 1	NT	NT	NT
Tumor-bearing + Queuine (0.05 A260 Units/hr)	2	77 ± 1	22 ± 1	0.2 ± 0.2	NT	NT	NT
Tumor-bearing	7	30 ± 10	53 ± 9	17 ± 4	80 ± 4	14 ± 3	6 ± 2
Tumor-bearing + Queuine (0.21 A260 Units/hr)	3 [†]	70 ± 4	28 ± 4	1.2 ± 0.4	89 ± 3	11 ± 3	0.6 ± 0.2
Control	4	-	-	-	83 ± 2	14 ± 1	3 ± 1

*Mice (male Swiss Albino, 30-40 grams) were inoculated with 10⁶ tumor cells on Day 0 and separated into three groups: (1) infused continuously with saline; (2) infused continuously with queuine; (3) received tumor alone. In Experiment 1, queuine (106 A260 units/ml in H₂O) or saline were delivered to the tumor-bearing animals with Alzet osmotic minipumps (Model 1702) at the rate of 0.5 l/hr; and animals were sacrificed on Day 9. In Experiment 2, queuine (214 A260 units/ml in H₂O) or saline were delivered with Alzet osmotic minipumps (Model 2001) at the rate of 1 ul/hr; and animals were sacrificed on Day 7. Pumps were implanted subcutaneously on Day 0. tRNA^{His} species were resolved as described in Fig. 2. The several different preparations of tRNA did not differ significantly with respect to amount of tRNA^{His} (data not shown). The data for tumor-bearing animals which received either saline or no treatment were grouped together because no difference was observed between them. Values are means ± standard deviations. NT signifies not tested.

[†]These tumor tRNA^{His} values are the means from only 2 of the 3 mice treated with queuine because in the third mouse the small amount of cells recovered (0.1 ml packed cell volume) and a significantly different pattern for tRNA^{His} (92% species 1; 8% species 2; 0.1% species 3) indicated that only normal peritoneal cells were detected.

previously to be [Q+] and [Q-], respectively (4,10,15), and the present and prior data (6,11) are consistent with $\text{tRNA}_{2}^{\text{His}}$ and $\text{tRNA}_{3}^{\text{His}}$ being Q-free precursors of $\text{tRNA}_{1}^{\text{His}}$. Table 1 shows that liver tRNA^{His} is 80-90% [Q+] and that tumor tRNA^{His} is less than 50% [Q+]. In two experiments, treatment of tumor-bearing animals with two different doses of queuine shifted the tumor tRNA^{His} species-distribution toward that observed for liver. These data (Table 1 and Fig. 2) are consistent with the incorporation of exogenous queuine into [Q-]tRNA in vivo.

It is of interest that four times more queuine was delivered in Experiment 2, than in Experiment 1, yet the tumor tRNA^{His} derived from both groups of treated mice was modified similarly. However, while no effect of queuine on tumor-growth was noted in Experiment 1 (data not shown), the higher dosage of queuine in Experiment 2 produced an apparent antitumor effect (Table 2). The total tumor recovered from the queuine-treated mice in Experiment 2 was less than that recovered from the untreated controls; indeed, no tumor was recovered from one animal. Moreover, the 7-day increase in body weight was prevented as well. Together these observations reflect impaired tumor-growth.

There are several possible explanations for the relative deficiency of Q in the tRNA of tumors, including: a) a defect in the biosynthesis, cell uptake, or insertion of precursor base (queuine or prequeuine) into tRNA; b) excessive catabolism of precursors of Q or removal of Q from tRNA; and c) a

TABLE 2. Effect of Queuine on the Growth of Ehrlich Ascites Tumor Cells in Mice*

Group	No. of Mice	Weight Ratio Days 7/0	Packed Volume of Tumor Cells (ml)
Tumor-bearing	7	1.29 ± .08	1.9 ± 0.6
Tumor-bearing + Queuine	3	1.10 ± .07	0.5 ± 0.5
Control	3	1.00 ± .05	--

*The tumor-bearing groups and nontumor-bearing control group from Experiment 2, Table 1, were weighed daily. One mouse from the control group in Table 1 was not weighed on Day 0 and is not represented here. The values represent the mean ± S.D. for each group.

tumor-elicited metabolite that interferes at some point in the normal metabolism of Q. With regard to these possibilities, tumor tissue has been reported to contain adequate levels of tRNA-guanine transglycosylase (7); however, the de novo synthesis of Q has yet to be demonstrated in mammals and nothing is known about the catabolism of Q. Additionally, the data in Table 1 allow the suggestion that the tumor may alter the metabolism of Q in non-tumorous tissues: a) tRNA^{His}₃ increased to a small extent in the livers of the tumor-bearing animals and b) treatment of these animals with queuine led to a decrease in liver tRNA^{His}₃ to below the values for control liver.

Finally, it is of interest that deficiencies of Q in tRNA, similar to those observed here in Ehrlich ascites tumors, have been observed in the livers of germ-free mice fed a defined diet (16). If otherwise apparently normal germ-free mice can be deficient in Q, the undermodification per se cannot be a sufficient requirement for tumorigenicity. The data at hand, however, are consistent with the possibility that a deficiency of Q favors tumor growth; therefore, the phenomenon reported here has potential significance for the therapy of neoplasia.

ACKNOWLEDGMENTS

We thank Dr. Walter Farkas for sharing a manuscript prior to publication and Peg Westmoreland, Mimi Ward and Mary Anne Ouellette for technical assistance. This work was supported by NIH grant CA202919, the University of Tennessee College of Medicine Research Contingency Fund, (JRK); Program Grant CA23099, Cancer Center (CORE) Grant CA21765, and by ALSAC (WTB).

REFERENCES

1. Kasai, J., Ohashi, Z., Harada, F., Mishimura, S., Oppenheimer, N.J., Crain, P.F., Liehr, J.G., von Minden, D.L. and McCloskey, J.A. (1975) *Biochemistry* **14**, 4198-4208.
2. Brisco, W.T., Griffin, A.C., McBride, C., and Bowen, J.M. (1975) *Cancer Res.* **35**, 2586-2593.
3. Olsen, C.E. and Penhoet, E.E. (1976) *Biochemistry* **15**, 4649-4654.
4. McNamara, A.L. and Smith, D.W.E. (1978) *J. Biol. Chem.* **253**, 5964-5970.
5. Yokoyama, S., Miyazawa, T., Titaka, Y., Yamaizumi, Z., Kasai, H., and Nishimura, S. (1979) *Nature* **282**, 107-109.
6. Katze J.R. and Farkas, W.R. (1979) *Proc. Natl. Acad. Sci. USA* **76**, 3271-3275.
7. Shindo-Okada, N., Okada, N. Ohgi, T., Goto, T., and Nishimura, S. (1980) *Biochemistry* **19**, 395-400.

8. Crain, P.F., Sethi, S.K., Katze, J.R. and McCloskey, J.A. (1980) *J. Biol. Chem.* (in press).
9. Okada, N., Shindo-Okada, N., Sato, S., Itoh, Y.H., Oda, K.-I, and Nishimura, S. (1978) *Proc. Natl. Acad. Sci. USA* 75, 4247-4251.
10. Katze, J.R. (1978) *Nucleic Acids Res.* 5, 2513-2524.
11. Roe, B.A., Stankiewicz, A.F., Rizi, H.L., Weisz, C., Dilauro, M.N., Pike, D. Chen, C.V. and Chen, E.Y. (1979) *Nucleic Acids Res.* 6, 673-688.
12. Marini, M. and Mushinsky, J.F. (1979) *Biochim. Biophys. Acta* 562, 252-270.
13. Landin, R.-M., Boissard, M. and Petrisant, G. (1979) *Nucleic Acids Res.* 7, 1635-1648.
14. Jackson, C.D., Irving, C.C., & Sells, B.H. (1970) *Biochim. Biophys. Acta.* 217, 64-71.
15. DuBrul E.F. and Farkas, W.R. (1976) *Biochim Biophys. Acta* 442, 379-390.
16. Farkas, W.R. *J. Biol. Chem.* (in press).
17. Okada, N., Harada, F. and Nishimura, S. (1976) *Nucleic Acids Res.* 3, 2593-2063.
18. White, B.N. (1974) *Biochim. Biophys. Acta* 353, 283-291.